Acute Toxicity of Dichlorvos (DDVP) to Fingerling Mirror Carp, *Cyprinus carpio* L.

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The increasing use of synthetic chemical pesticides is causing worldwide pollution. Although pesticides by their nature are toxic and are designed to kill unwanted organisms, they, when applied to the land, may be washed into surface waters and kill or at least adversely influence the life of aquatic organisms (Hayes and Laws 1991; Rand 1995). Because of high acute toxicity and the consequent dangers to workers, there are concerns whether safe use of pesticides is possible under such conditions. However, dichlorvos (DDVP) still is widely used in developing countries such as Turkey to control of pests.

The environmental fate and effects of DDVP have been well-summarized (EPA 2000). DDVP is generally found to be toxic to phytoplankton (EPA 2000; Raine et al. 1990), zooplankton (EPA 2000; Sanchez-Fortun et al. 1995), molluscs (Jonnalagadda and Rao 1996; Tripathi and Agarwal 1998), crustaceans (Geraldine et al. 1999; McHenery et al. 1996) and fish (Chuiko and Slynko 1995; Dutt and Guha 1988; Eisler 1970; EPA 2000; Mayer and Ellersieck 1986; Verma et al. 1981). The effects of DDVP on reproduction (Benarji and Rajebdranath 1991; Saksena and Saxena 1999), enzyme activity (Golovanova et al. 1994; Pavlov et al. 1992), respiration (Mohapatra and Noble 1993), genetics (Rishi and Grewal 1995), feeding behavior (Pavlov et al. 1992), general biochemistry (Saxena and Saksena 1996) and hematological systems (Rajeswari et al. 1989) of fishes were also reported.

In Turkey, larvae of some commercial warm-water fish species, such as carp are artificially reared and then introduced to freshwater reservoirs when they have reached the fingerling stage (approx. 5-10 cm in length). At the same time, dichlorvos (0,0 dimethyl-2,2-dichlorovinyl phosphate; DDVP) is a commonly used pesticide for pest control in the agricultural fields around these freshwater reservoirs. Therefore, the present study has aimed to determine the acute toxicity of DDVP on fingerling mirror carp at the size attained just before introduced into reservoirs. Our findings would be a useful tool for the effective management and control of regional reservoirs with respect to the imput of DDVP from agricultural areas.

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MATERIALS AND METHODS

In this study, a static acute toxicity bioassay was performed according to standard methods (APHA 1999) to determine the 24, 48, 72 and 96 h lethal concentrations of dichlorvos (0,0-dimethyl-2,2-dichlorovinyl phosphate; DDVP) for fingerling mirror carp. Experimental animals from 3.5-4.01 (3.72±0.20) g and 6.4-7.1 (6.8±0.3) cm in length, were obtained from Keban Fish Breeding Unit of DSI Ninth Region Directorate, Elazig, Turkey. They were brought to the laboratory at Keban Professional School and acclimatized for 4 days under standard laboratory conditions. Fishes were fed with flour pellets during adaptation. However, they were not fed during the last 24 h of adaptation and throughout the duration of the test. Before the starting of the test, all experimental aquaria (60 L capacity) were cleaned and filled with 50 L of dechlorinated tap water (temperature: 20.6 °C±0.3, pH: 8.45±0.03, dissolved oxygen: 7.17±0.25 mg L⁻¹, electrical conductivity: 232.25±10.54 μS cm⁻¹, alkalinity: 142-168 mg L⁻¹ and hardness: 186-240 mg L⁻¹ as CaCO₃). Five different concentrations (4, 8, 16, 32, 64 µg L⁻¹) of DDVP. with three replicates, were used in this test. Control units with three replicates were also prepared. Aeration was applied to the aquaria for 2 h in order to obtain a homogeneous concentration of the toxic compound, and then 10 fish were transferred in each test aquarium. Mortality was assessed at 24, 48, 72 and 96 h after the start of the tests. Dead animals were removed immediately. Behavioral changes of test animals were closely followed and recorded. Toxicity was assigned based on LC₅₀, according to the guidelines given by Kamrin (1997). DDVP is classified as slightly toxic to moderately toxic for fish, with an LC₅₀>1 mg L⁻¹.

The results were expressed as mean \pm standard deviation (SD) or range. All replicates were used for calculation of mean values. Statistical analysis was performed with the SPSS statistical software package (Version 10.1, SPSS Inc., Chicago, Illinois, USA). The LC₅₀ and 95% confidence limits were calculated by the probit method. Duncan's multiple range tests was employed for comparing the significance level for time using a significance level of 0.05.

RESULTS AND DISCUSSION

The behavioral responses of the test animals were observed at 1-10 h during the first day of exposure and then every 12 h during the last three days of exposure. Normal behavior was observed for control animals and those at 4 mg L⁻¹ concentration. Initial changes in behavioral response were observed 1 h after exposure in the highest DDVP concentrations (64 mg L⁻¹). Animals exposed to 8 and 16 mg L⁻¹ concentrations showed normal behavior until the end of the first day of exposure. However, afterward they showed some abnormal behavior such as reduced general activity and loss of equilibrium when compared with the control animals. The two highest concentrations (32 and 64 mg L⁻¹) showed all behavioral responses at high levels and the onset was within the first 1 h after starting the test. Abnormal behavioral responses observed at all concentrations above 16 mg L⁻¹ were rapid gills movement, erratic swimming, swimming at the

water surface, air gulping at the water surface and staying motionless on the aquarium bottom later in exposure.

The calculated 24, 48, 72 and 96h LC₅₀ values (95% confidence limits) for DDVP, using a static bioassay system to fingerling mirror carp were 84.99 (69.89-138.95), 67.86 (48.67-127.75), 26.18 (20.44-35.06) and 9.41 (7.54-11.49) mg L⁻¹ respectively (Table 1). The results show that DDVP was slightly toxic at 24, 48 and 72 h and moderately toxic at 96 h to fingerling mirror carp. The toxicity of this compound on experiment animals increased with increasing concentration and exposure time. There were significant differences between LC50 values calculated for 24, 48, 72 and 96 hours (p<0.05). All animals exposed to 64 mg L⁻¹ of DDVP were dead after 96 h of exposure (Table 1). The percentage of living animals in 32 mg L⁻¹ concentration was only 10% at the end of the test. It was calculated that when the concentration of DDVP increased approximately 5.6 times, the percentage of dead animals increased by 50 %. The regression equation y = -0.309 + 0.381Log(x) with a correlation coefficient of 0.976 was obtained by probit method. Where; x and y denote DDVP concentration and the corresponding probit value, respectively. This equation can provide to calculate predicted toxicity at any other concentrations of DDVP. The calculated safe concentration of DDVP for fingerling mirror carp was 94.1 µg L⁻¹ (i.e., 1/100 of LC₅₀ value).

Table 1. Cumulative mortality and LC₅₀ values with 95% confidence limits (in parentheses) of DDVP for fingerling mirror carp.

| No. of us fish — | Exposure time (hours) | | | | | |
|------------------|--|--|---|---|--|--|
| Exposed | 24 | 48 | 72 | 96 | | |
| 30 | 6 | 15 | 24 | 30 | | |
| 30 | 1 | 6 | 17 | 27 | | |
| 30 | 0 | 2 | 9 | 24 | | |
| 30 | 0 | 1 | 6 | 15 | | |
| 30 | 0 | 0 | 0 | 2 | | |
| 30 | 0 | 0 | 0 | 0 | | |
| | 30 30 30 30 30 30 30 | 30 6 30 1 30 0 30 0 30 0 30 0 | 30 6 15 30 1 6 30 0 2 30 0 1 30 0 0 | 30 6 15 24 30 1 6 17 30 0 2 9 30 0 1 6 30 0 0 0 0 | | |

LC₅₀ values, mg L⁻¹ 84.99^a 67.86^b 26.18^c 9.41^d (95 % confidence limits)(69.89-138.95)(48.67-127.75)(20.44-35.06) (7.54-11.49)

LC₅₀ values in row with different letters significantly differ (p<0.05)

Some pesticides used in agriculture are known to be toxic to non-target aquatic biota; DDVP belongs to this group. The toxic effects of DDVP to various fish species are summarized (Table 2). In general, the toxic effect of DDVP changes with respect to species and size of animals and the duration of exposure. The results of the present study showed that the toxicity of DDVP to fingerling mirror carp under the conditions used, was also time dependent.

Table 2. Toxicity studies for DDVP on various fish species.

| Scientific Name | Fish | Expo- | LC50, | Chemical | Experi- | Acute | References |
|------------------------|--------|--------------|--------------|-----------|---------------|----------|--------------------|
| | Size | sure | mg L | Descrip- | ment | Toxicity | |
| | | Time | | tion | Туре | Rating | |
| Abramis brama | NR | 24 h | | 97.5 % | Renewal | ST | Chuiko and |
| | | 48 h | | AI, DDVP | • | | Slynko 1995 |
| | | 72 h | 21.11 | | | | |
| | 5.0 | 96 h | 16.66 | ND | 04-4 | MT | E'-1 1070 |
| Anguilla rostrata | 5.9cm | 24 h | 2.3 2.3 | NR | Static | MT | Eisler 1970 |
| | 0.14g | 48 h 96 h | 1.8 | | | | |
| Onhiceanhalus nimetata | 0.11am | | 4.3 | EC 76 % | Static | MT | Verma et al. |
| Ophiocephalus punctata | 40-55g | | 3.4 | EC /0 70 | Static | 141 1 | 1981 |
| | | 72 h | 2.1 | | | | 1701 |
| | | 96 h | 2.3 | | | | |
| Cyprinus carpio | 4.5- | 24 h | 9.44 | DDVP | Static | MT | Dutt and |
| Cypi www our pro | 5.5cm | 48 h | 8.99 | | | | Guha 1988 |
| | 1.1- | 72 h | 8.21 | | | | |
| | 1.4g | | | | | | |
| Fundulus heteroclitus | 5.5cm | 24 h | 3.41 | NR | Static | MT | Eisler 1970 |
| | 1.7g | 48 h | 2.68 | | | | |
| | | 96 h | 2.68 | | | | |
| Fundulus heteroclitus | 4.0g | 96 h | 3.2 | NR | Static | MT | EPA 2000 |
| Fundulus majalis | 0.9g | 96 h | 2.3 | | | MT | |
| Gambusia affinis | 0.2g | 24 h | 17.8 | Technical | Static | ST | Mayer and |
| | | 96 h | 5.27 | Material | | MT | Ellersieck 1986 |
| Lepomis macrochirus | 1.5g | 24 h | 3.08 | Technical | Static | MT | |
| | | 96 h | 0.87 | Material | | HT | |
| Lepomis macrochirus | 0.09g | 96 h | 4.30 | NR | Flow | MT | EPA 2000 |
| | 0.4g | 96 h | 0.18 | NR | Static | HT | |
| | 0.8g | 96 h | 0.80 | NR | Static | HT | |
| Menidia menidia | 2.1g | 24 h | 9.60 | NR | Static | MT | EPA 2000 |
| Mugil cephalus | 8.4cm | 24 h | 0.74 | NR | Static | HT | Eisler 1970 |
| | 6.4g | 48 h | 0.66 | | | HT | |
| | 4.6cm | 96 h 24 h | 0.20 1.75 | | | HT MT | |
| | 1.0g | 48 h | 1.75 | | | MT | |
| | 1,05 | 96 h | 0.25 | | | HT | |
| Mystus vittatus | 8-10cm | | 0.73 | EC 76% | Static | HT | Verma et al. |
| | 6-10g | 48 h | 0.65 | | ~~ >40 > 1 40 | HT | 1981 |
| | | 72 h | 0.51 | | | HT | |
| | | 96 h | 0.45 | | | HT | |
| Oncorhynchus clarki | 0.17g | 96 h | 0.213 | NR | Static | HT | EPA 2000 |
| Oncorhynchus mykiss | 2.3g | 96 h | 0.100 | NR | Static | HT | EPA 2000 |

Narrative descriptions of toxicity were assigned based on LC₅₀ according to the guidelines in Kamrin (1997). ST, Slightly Toxic (LC₅₀ = 10-100 mg L⁻¹); MT, Moderately Toxic (LC₅₀ =1-10 mg L⁻¹); HT, Highly Toxic (LC₅₀ = 0.1-1 mg L⁻¹); VHT, Very Highly Toxic (LC₅₀ < 0.1 mg L⁻¹); NR, Not Reported; EC, Effective Concentration; AI, Active Ingredient

In laboratory tests, the USDA National Agricultural Pesticide Impact Assessment Program's EXTOXNET document reports dichlorvos acute toxicity to fish at 96 h of 11.6 mg L⁻¹ in fathead minnow, 0.9 mg L⁻¹ in bluegill, 5.3 mg L⁻¹ in mosquito fish and 1.8 mg L⁻¹ in American eels. Pesticide Action Network (PAN) pesticides database also reports DDVP average acute toxicity for fish at 96 h of 1-10 mg L⁻¹. Our results are in good agreement with these two reports. Similar ranges of DDVP LC₅₀ values for various fish species were also reported in some studies (Table 2).

REFERENCES

- APHA (1999) Standard methods for the examination of water and wastewater. APHA- AWWA-WEF, Washington, DC
- Benarji G, Rajebdranath T (1991) Dichlorvos-induced histoarchitectural changes in the oocytes of a freshwater fish. Funct Dev Morphol 1: 9-12
- Chuiko GM, Slynko YV (1995) Relation of allozyme genotype to survivorship of juvenile bream, *Abramis brama* L., acutely exposed to DDVP, an organophosphorus pesticide. Bull Environ Contam Toxicol 55: 738-745
- Dutt N, Guha RS (1988) Toxicity of a few organophosphorus insecticides to fingerlings of bound water fishes, *Cyprinus carpio* L. and *Tilapia mossambica* Peters. Indian J Entomol 50: 403-421
- Eisler R (1970) Acute toxicities of organochlorine and organophosphorus insecticides to estuarine fishes. Technical Paper No 46, USDI Bureau of Sport Fish and Wildlife, Washington DC
- EPA (2000) Environmental Protection Agency, Office of Pesticide Programs, Pesticide Ecotoxicity Database, Environmental Fate and Effects Division, USEPA, Washington, DC 2000
- Geraldine P, Bhavan PS, Kaliamurthy J, Zayapragassarazan Z (1999) Effects of dichlorvos intoxication in the freshwater prawn, *Macrobrachium malcolmsonii*. J Environ Biol 20: 141-148
- Golovanova IL, Chuiko GM, Pavlov DF (1994) Effects of cadmium, naphthalene and DDVP on gut carbohydrases activity in bream (*Abramis brama* L.) and Mozambique tilapia (*Oreochromis mossambicus* Peters). Bull Environ Contam Toxicol 52: 338-345
- Hayes WJ, Laws ER (1991) Handbook of pesticide toxicology (3 vols). Academic Press, San Diego
- Jonnalagadda PR, Rao BP (1996) Histopathological changes induced by specific pesticides on some tissues of the fresh water snail, *Bellamya dissimilis* Muller. Bull Environ Contam Toxicol 57: 648-654
- Kamrin AC (1997) Pesticide profiles: Toxicity, environmental impact, and fate. Lewis Publishers Inc, Boca Rotan Florida
- Mayer FL, Ellersieck MR (1986) Manual of acute toxicity: Interpretation and data base for 410 chemicals and 66 species of freshwater animals. US Department of the Interior, Fish and Wildlife Services. Resource Publication 160, Washington DC
- McHenery JG, Francis C, Davies IM (1996) Threshold toxicity and repeated exposure studies of dichlorvos to the larvae of the common lobster (*Homarus gammarus* L.). Aquat Toxicol 34: 237-251

- Mohapatra BC, Noble A (1993) changes in oxygen uptake in mullet, *Liza parsia* (Hamilton-Buchanan) exposed to dichlorvos. Fish Technol 30: 112-114
- Pavlov DD, Chuiko GM, Gerassimov YV, Tonkopiy VD (1992) Feeding behavior and brain acetylcholinesterase activity in bream (*Abramis brama* L.) as affected by DDVP, an organophosphorus insecticide. Comp Biochem Physiol C103: 563-568
- Raine RCT, Cooney JJ, Coughlan MF, Patching JW (1990) Toxicity of nuvan and dichlorvos towards marine phytoplankton. Bot Mar 33: 533-537
- Rajeswari K, Reddy SJ, Reddy DC, Ramamurthi R (1989) Effect of dichlorvos on certain blood parameters of the fish *Clarias batrachus*. Environ Ecol 7: 933-934
- Rand GM (1995) Fundamentals of aquatic toxicology. Taylor and Francis Publishing, Washington
- Rishi KK, Grewal S (1995) Chromosome aberration test for the insecticide, dichlorvos on fish chromosomes. Mutat Res 344: 1-4
- Saksena DN, Saxena V (1999) Organophosphorus pesticides induced changes in the ovarian activity of a freshwater murrel, *Channa orientalis* (Schneider): A histological study. J Environ Biol 20: 157-162
- Sanchez-Fortun S, Sanz-Barrera F, Barahona-Gomariz MV (1995) Acute toxicities of selected insecticides to the aquatic arthropod, *Artemia salina*. Bull Environ Contam Toxicol 54: 76-82
- Saxena V, Saksena DN (1996) Effect of nuvan and dimecron on the ovarian biochemical constituents in a freshwater murrel, *Channa orientalis* (Schneider). J Environ Pollut 3: 157-164
- Tripathi AM, Agarwal RA (1998) Molluscicidal and anti-AChE activity of tertiary mixtures of pesticides, Arch Environ Contam Toxicol 34: 271-274
- Verma SR, Rani S, Bansal SK, Dalela RC (1981) Evaluation of the comparative toxicity of thiotox, dichlorvos and carbofuran to two freshwater teleosts *Ophiocephalus punctatus* and *Mystus vittatus*. Acta Hydrochim Hydrobiol 9: 119-129